ORIGINAL ARTICLE



In-Silico Study on Anti-bacteria and Anti-fungal Activities of 3,4-Dihydropyrimidin-2(1*H*)-One Urea Derivatives

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Abstract

The menace caused by fungi amidst human being and in the world at large is colossal. Their increasing rate of causing havoc has drawn the attention of scientist to developing efficient drug-molecules in order to cure these diseases. In this work, sixteen molecular compounds were studied with aim of investigating the non-bonding interaction between the studied compounds and the receptors using the following software Spartan 14 (optimization), Pymol (for treating downloaded protein), Autodock Tool (for locating binding site in the downloaded protein and for converting ligand and receptor to.pdbqt format from.pdb format), Auto dock vina (for docking calculation) and discovery studio (for viewing the non-bonding interaction between the docked complexes) so as to observe anti-*e-coli* and anti-1,4 α -demethylase properties of 3,4-dihydropyrimidin-2(1*H*)-one urea derivatives. The calculated parameters such as E_{HOMO} energy, E_{LUMO} energy, Dipole moment, log P, HBD and HBA which described anti-*e-coli* and anti-1,4 α -demethylase portential anti-2(1*H*)-one Urea Derivatives were observed. This showed that all the studied compound **4** and **3** possess better ability to inhibit *e-coli* and 1,4 α -demethylase better than other studied compounds. More so, ADMET properties of five selected compounds were determined and it was observed that the ADMET properties obtained for the selected compounds were similar to the ADMET properties of the standards used in this work.

Keywords In-silico · Fungi · 3,4-dihydropyrimidin-2(1H)-one urea · DFT · Docking

1 Introduction

The continuous design and development of drugs has reduced mortality and morbidity rate globally. Growing resistance by fungi and bacteria to drugs has led to incessant and increasing rate of death [1]. Many researchers have reported that several antibiotics are becoming ineffective and this has prompted scientists all over the world to searching out lasting solution via medicinal plant and synthesized compounds to combat this menace [2]. Therefore, the need to design and develop efficient anti-fungi and anti-bacterial drugs has drawn the attention of scientist over the years [3]. Bacteria are numerous organisms which usually exist among human being [4–6]. They are composed of great area of prokaryotic microorganisms. They are very small in size and they cannot be seen except it is placed under lenses [7]. Bacteria have series of shapes (Sphere, rod and spiral) [8]. As reported by Georgiana et al. [9] some bacteria strains are pathogenic in nature and they have been reported to be responsible for several infections such as urinary tract infection, biliary tract infection, wound infections etc [10, 11]. Infections from bacteria have been regarded has one of the frontline cause of death globally and due to this, it is important that patients with infections from bacterial receives urgent treatment [12].

 14α -demethylase is a vital protein for ergosterol's biosynthesis in fungi and it is a major target for drug-like molecules [13]. It is a vital enzyme that helps in converting lanosterol to lower molecules. It exists in several organisms and most especially in fungi where it acts as an intermediary in membrane absorptivity [14]. Fungal infection has also become cause of death among immune compromised human being.

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As reported by many researchers, several drug-like compounds such as voriconazole, echinocandins, ketoconazole, amphotericin B and nikkomycin have been used to combat fungi activities in human [15]. Despite all these efforts put in place by scientists to end fungi activities in human, the rate at which it operates keep increasing [16, 17]. According to Warrilow et al. [18], Sterol 1,4- α -demethylase was classified to be cytochrome P450 superfamily and it was observed to be one of the vital agent to synthesized ergosterol fungi. It also acts as an agent that biologically synthesized cholesterol in mammals.

Dihydropyrimidine derivatives are crucial molecular compounds in medicinal world. Their pharmacological and biological features have greatly drawn the attention of several scientists globally. Dihydropyrimidine derivatives could be used as antiviral, antibacterial, antifungal, antihypertensive and anticancer agents [19-22]. 3,4-Dihydropyrimidin-2-(1H)-one was first synthesized using Biginelli reaction and this was first accomplished in 1893 and several biological properties were reported by many researchers [23, 24]. Also, several scientists reported that N1-substituted Dihydropyrimidine derivatives possess more improved pharmacological activity [25-27]. In this work, sixteen molecular compounds were theoretically studied with the aim of observing the molecular descriptors responsible for anti- E-coli and anti-1,4 α -demethylase activities of 3,4-dihydropyrimidin-2(1H)-one Urea Derivatives and developing efficient QSAR model to predict the biological activities of the studied compounds as well as observing biological interaction present between the studied compounds and E coli (PDB ID: 5r1r) [28] as well as 1,4 α -demethylase (PDB ID: 3juv) [29] via docking study.

2 Materials and Methods

2.1 Optimization, QSAR and Docking Studies

In this present study, 2D structure of sixteen molecular compounds (Table 1) were accomplished using CHEMDRAW ultra 8.0 version and were optimized using density functional theory with 6-31G* (B3LYP) as basis set via Spartan'14 [30–34]. The studied compounds were optimized so as to generate descriptors that describe anti-1,4 α -demethylase and anti-bacteria activities. In this work, series of molecular descriptors were calculated and eleven electronic descriptors were selected and were used for QSAR study. Also, quantitative structural activities relationship (QSAR) models (Eq. 1 for *Ecoli* and Eq. 2 for 1,4 α -demethylase) were developed using selected electronic descriptors using Gretl software so as to predict the observed bioactivity of the studied compounds. The observed IC₅₀ was used as dependent variable and the calculated descriptors were used as independent
 Table 1
 Schematic structure of 3,4-dihydropyrimidin-2(1H)-one urea derivatives

	S
1	2-F
2	2-Cl
3	2-CF ₃
4	2-OC ₆ H ₅
5	2-F, 6-CH ₃
6	2-F, 6-CF ₃
7	2-Cl, 6-CH ₃
8	2-Cl, 6-CF ₃
9	2-Cl, 5-CF ₃
10	2-Cl, 4-CF ₃
11	2-Cl, 6-F
12	3-CF ₃
13	3-Cl, 4-F
14	4-isopropyl
15	4-CF ₃
16	4-OCH ₃

variables and the descriptors used were carefully selected so as to avoid multi-collinearity. Correlation coefficient (R^2) , adjusted R^2 , P value, mean square error (MSE) were observed in developing the QSAR model.

The optimised molecular compounds were subjected to docking to calculate binding affinity for individual studied compound and observe the non-bonding interaction existing between the complexes. The software used were Pymol (for treating downloaded protein by removing other foreign compounds downloaded together with the protein), Autodock Tool (for locating binding site in the downloaded protein and for converting ligand and receptor to.pdbqt format from.pdb format), Auto dock vina (for docking calculation) and discovery studio (for viewing the non-bonding interaction between the docked complexes). The grid box (dimension and centre) were: center (X = -5.942, Y = 94.018,Z = 24.16) and size (X = 98, Y = 90, Z = 74) for *E coli* (PDB) ID: 5r1r): and center (X = -82.519, Y = 25.442, Z = -5.492) and size (X = 74, Y = 58, Z = 96) for fungi (PDB ID: 3juv). The spacing was set to be 1.00 Å.

$IC_{50} = -3695.81 - 22.3862(E_{HOMO}) + 29.0752(E_{LUMO}) - 1$ - 7.21692(Vol) - 0.0737399(PSA) + 89.4820(Pol) - 2.2	- (1)
$R^2 = 0.821$, $AdjR^2 = 0.665$, $P < 0.0001$, F value = 5.25, $MSE = 0.019862$	orbital energy (E_{HOMO}), lowest unoccupied molecular orbital energy (E_{LUMO}), polarizability, dipole moment, hydrogen bond acceptor, energy band gap, area, volume, polar sur-
$IC_{50} = -2020.78 - 13.5155(E_{HOMO}) + 17.4543(E_{LUMO}) - 1$ $- 3.92066(Vol) + 48.7551(Pol) - 1.90451(NOR)$.44899(LogP) (2)

 $R^2 = 786$, $AdjR^2 = 0.643$, P < 0.0001, F value = 5.514, MSE = 0.020969

2.2 ADMET Properties

Phsyicochemical and Absorption, Distribution, Metabolism, Excretion and the Toxicity properties of the studied molecular compounds were studied via admetSAR (http://lmmd. ecust.edu.cn/admetsar1) [35]. Many properties were put in to consideration such as Human Intestinal Absorption (HIA), Blood Brain Barrier, Ames test, Caco-2 cell permeability.

3 Result and Discussion

3.1 Molecular Descriptors

 Table 2
 Calculated

 molecular parameters from
 3,4-dihydropyrimidin-2(1H)

 one
 Urea
 Derivatives

The calculated molecular descriptors obtained from optimized studied compounds are highest occupied molecular face area, Log P, hydrogen bond donor (Table 2). In this study, the role of dipole moment as one of the molecular non-bonded interaction in drug design cannot be over emphasized [35, 36]. As reported by Oyewole et al. [37], as well as Oyebamiji et al. [38], molecular compound with high dipole moment value is expected to have anomalous property. As shown in Table 2, all the studied compounds can act as potential and fungal and antibacterial agent.

Also, calculated E_{HOMO} was obtained from the optimized 3,4-dihydropyrimidin-2(1*H*)-one Urea derivatives so as to determine their ability to release electron to the neighbouring compound [39, 40]. As shown in Table 2, compound **16** possess better ability to release electron to the nearby compounds and it is expected to inhibit more than other studied compounds. Also, the calculated E_{LUMO} gotten from the studied molecules reveal their tendency to receive electron from the nearby molecules [41, 42]; thus, compound **13** possess higher ability to receive electron from nearby compounds than other studied ligands.

Mol	E _{HOMO}	E _{LUMO}	BG	MW	LogP	VOLUME	PSA	Pol	HBD	HBA	NOR
1	-5.76	-1.35	-4.41	428.49	2.32	411.06	71.73	73.68	4	7	3
2	-5.87	-1.4	-4.47	444.94	2.72	418.99	67.75	74.31	4	7	3
3	-5.86	-1.42	-4.44	478.5	3.08	437.19	66.84	75.79	4	7	3
4	-5.61	-1.36	-4.25	502.6	3.18	498.41	74.02	80.8	4	8	4
5	-5.78	-1.4	-4.38	442.52	2.8	428.41	69.13	75.09	4	7	3
6	-5.85	-1.41	-4.44	496.49	3.24	442.23	69.88	76.2	4	7	3
7	-5.79	-1.39	-4.4	458.97	3.2	437.74	69.38	75.84	4	7	3
8	-5.85	-1.36	-4.49	512.94	3.64	451.46	69.35	76.93	4	7	3
9	- 5.95	-1.4	-4.55	512.94	3.64	450.89	67.99	76.87	4	7	3
10	-5.85	-1.39	-4.46	512.94	3.64	452.18	71.65	77	4	7	3
11	-5.86	-1.4	-4.46	462.93	2.87	424.58	71.2	74.76	4	7	3
12	-5.83	-1.42	-4.41	478.5	3.08	437.74	69.02	75.84	4	7	3
13	-5.81	-1.43	-4.38	462.93	2.87	423.94	69.04	74.73	4	7	3
14	- 5.59	-1.4	-4.19	452.58	3.48	460.82	69.12	77.76	4	7	3
15	-5.85	-1.4	-4.45	478.5	3.09	437.54	68.92	75.82	4	7	3
16	-5.35	-1.41	-3.94	440.52	2.03	432.77	76.05	75.55	4	8	3

BG band gap, *MW* Molecular weight, *LogP* lipophilicity, *PSA* polar surface area, *HBD* hydrogen bond donor, *HBA* hydrogen bond acceptor, *NOR* number of rings

Other descriptors obtained are Molecular Weight, Hydrogen Bond Donor, Hydrogen Bond Acceptor and Log P; all these descriptors were found to fall within the standard range (molecular weight \leq 500, hydrogen bond donor \leq 5, Hydrogen bond acceptor \leq 10 and Log P \leq 5) except compound **4**, **8**, **9** and **10** with higher molecular weight. This showed that all the studied compounds possess drug potential.

3.2 QSAR Study

The developed QSAR models (Eqs. 1 and 2) which comprises of selected descriptors were used to predict observed inhibition concentration (IC₅₀) for Ecoli and 14 α -demethylase. E_{HOMO}, E_{LUMO}, LogP, Vol, PSA, Pol, NOR for ECOLI and E_{HOMO}, E_{LUMO}, LogP, Vol, Pol, NOR $(14\alpha$ -demethylase) have played a serious role in predicting biological activities of 3,4-dihydropyrimidin-2(1H)one urea derivatives. As shown in Eq. 1, decreasing E_{HOMO}, LogP, Volume, PSA, and NOR as well as increasing E_{LUMO} and polrizability enhanced the anti-ecoli activity of,4-dihydropyrimidin-2(1H)-one Urea Derivatives. Also, decreasing E_{HOMO}, LogP, Volume, and NOR as well as increasing E_{LUMO} and polrizability improved the anti- 14α -demethylase activity of 4-dihydropyrimidin-2(1H)one Urea Derivatives (Eq. 2). As shown in Table 3, the predicted IC_{50} were closer to the observed IC_{50} , this show the efficiency of the developed QSAR model (Figs. 1 and 2). Also, it was observed that the lower the calculated residual, the closer the predicted IC_{50} to the experimental IC₅₀.

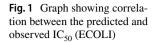
3.3 Molecular Docking Results

The optimized compounds were subjected to docking studies via discovery studio, autodock tool, autodock vina and pymol software. Docking studies was carried out on the studied compounds so as to observe the non-bonding interaction present between 3,4-dihydropyrimidin-2(1H)one urea derivatives and E coli (PDB ID: 5r1r) as well as $1,4\alpha$ -demethylase (PDB ID: 3juv). In order to ascertain the authenticity of the employed docking method, the docking method was validated by re-docking the native ligand into the active site of $1,4\alpha$ -demethylase (PDB ID: 3juv) so as to detect the likeness between the re-docked compound with best conformation to the posture of the native molecule (Fig. 3). Hence, the detected comparison as well as the root mean square deviation (RMSD) which occurs between the re-docked native molecule and the native ligand was closer to 1; therefore, this verified the reliability of the employed docking method.

All the studied compounds are very active against *E coli* and fungi as shown in Table 4. Compound **3** and **4** with -8.9 kcal/mol and -8.4 kcal/mol proved to be better than other studied compounds as anti-14 α -demethylase and anti*e-coli* respectively. As shown in Table 4, the standard compounds i.e. amoxylin and fluconazole have a lower binding affinity compared to the binding affinity obtained for each of the studied 3,4-dihydropyrimidin-2(1*H*)-one urea derivative. In this work, all the studied compounds possess potential ability to inhibit *E-coli* and 1,4 α -demethylase than the standard used and as shown in Figs. 4 and 5 urea parts of the studied compounds showed no interaction with the studied protein. More so, the residues involve in the interaction between the studied compounds and the *E coli* (PDB

Predicted (IC ₅₀)		E. coli	Predicted IC ₅₀	Residual	14α -demethylase	Predicted IC ₅₀	Residual
thylase	1	5.0132	5.0206	-0.0074	5.0132	5.0836	-0.0704
	2	5.0132	4.9014	0.1118	5.0132	4.7429	0.2703
	3	4.8239	4.7401	0.0838	4.6020	4.5386	0.0634
	4	4.4559	4.4559	0.0000	4.3979	4.3979	0.0000
	5	4.5228	4.4845	0.0383	4.3010	4.5069	-0.2059
	6	4.3979	4.6713	-0.2734	4.2596	4.5756	-0.3160
	7	4.2596	4.1934	0.0662	4.3976	4.2236	0.1740
	8	4.2218	4.3089	-0.0871	4.2218	4.2723	-0.0505
	9	4.0969	4.2295	-0.1326	4.2218	4.2351	-0.0133
	10	4.6020	4.3346	0.2674	4.4559	4.3386	0.1173
	11	4.0705	4.1356	-0.0651	4.3979	4.4137	-0.0158
	12	4.6989	4.4126	0.2863	4.6020	4.4145	0.1875
	13	4.0705	4.2377	-0.1672	4.0969	4.2609	-0.1640
	14	4.2596	4.2881	-0.0285	4.0457	4.0613	-0.0156
	15	5.0132	5.0888	-0.0756	4.8239	4.8284	-0.0045
	16	4.8239	4.8408	-0.0169	5.0132	4.9698	0.0434

Table 3 Observed and Predictedinhibition concentration (IC $_{50}$)for Ecoil and 14 α -demethylase



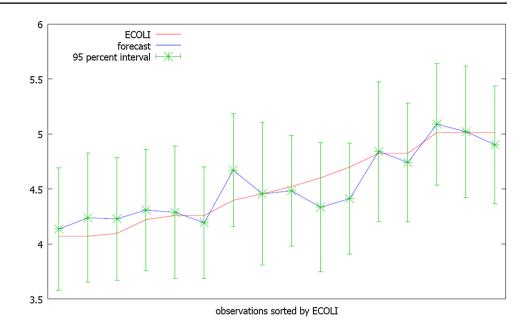
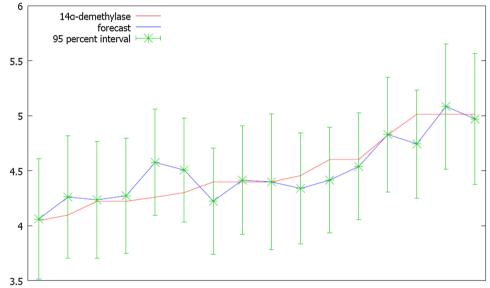


Fig. 2 Graph showing correlation between the predicted and observed IC_{50} (14 α -demethylase)

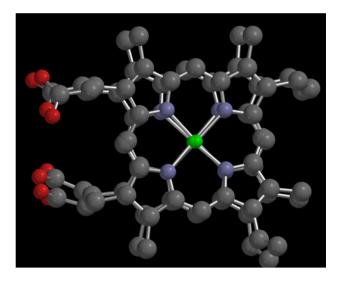


ID: 5r1r) as well as 1,4 α -demethylase (PDB ID: 3juv) were displayed in Figs. 4 and 5.

3.4 ADMET Properties Prediction of selected Compounds that Describe Anti-e-coli Activities of 3,4-Dihydropyrimidin-2(1H)-One Urea Derivatives

Table 4 Show the ADMET properties of selected compounds with more efficient binding affinity than other studied compounds as well as amoxicillin (**3**, **4**, **8**, **9** and **12**) via admetSAR server. According to Khaled et al., [43], it was reported that drug-molecules that have higher human observations sorted by 14a-demethylase

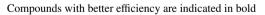
intestinal absorption value tends to absorb well. Thus, as displayed in Table 4, the value obtain for human intestinal absorption (HIA) for all the studied compounds fell within the same range and they were lower compare to the value obtain for amoxicillin (Standard). Also, the blood brain barrier value for the selected compounds fall within the same ranges shown in Table 4. The observed P-glycoprotein (nonsubstrate and non-inhibitor) reported for all the selected molecules were similar to the amoxicilin. AMES was observed so as to determine the level of mutagenicity; thus, all the



 $\ensuremath{\mbox{Fig. 3}}$ Overlay of native drug-like compounds over re-docked drug compound



	Binding affinity (<i>E coli</i>) (kcal/mol)	Binding Affinity (1,4α-demethylase) (kcal/mol)
1	-7.2	-7.5
2	-7.0	-7.5
3	-7.4	-8.9
4	-8.4	-7.2
5	-6.5	-6.9
6	-7.3	-7.7
7	-6.8	-7.5
8	-7.5	-7.7
9	-7.8	-7.1
10	-6.9	-8.6
11	-7.0	-7.5
12	-7.5	-7.4
13	-7.1	-7.5
14	-6.5	-7.4
15	-7.2	-7.9
16	-6.8	-7.6
Amoxicillin	-6.5	-
Fluconazole	_	-6.5



selected compounds were negative and this correlated with the standard (Amoxicilin). More so, the selected compounds were not carcinogenic and it also agreed with the standard (Table 5). The calculated cytochrome P450 showed that it could be inhibited by some of the studied compounds.

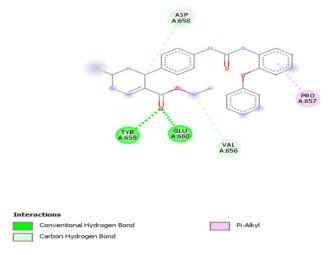


Fig. 4 Molecular interaction of compound 4 with 5r1r

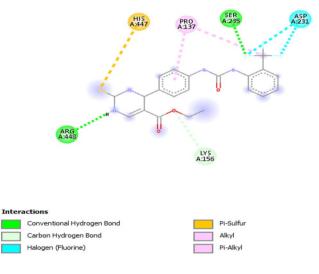


Fig. 5 Molecular interaction of compound 3 with 3 juv

3.5 ADMET Properties Prediction of selected Compounds that Describe Anti-1,4α-Demethylase Activities of 3,4-Dihydropyrimidin-2(1*H*)-One Urea Derivatives

ADMET properties of best five (5) studied compounds (3, 6, 8, 10 and 15) with potential anti-1,4 α -demethylase properties were considered. As shown in Table 5, HIA value obtained for the selected compounds were very closer to that of Fluconazole. AMES was observed so as to determine the level of mutagenicity; thus, all the selected compounds were negative and this correlated with the standard

Mode	Compound 3		Compound 4		Compound 8		Compound 9		Compound 12		Amoxylin	
	Result	Probability	Result	Probability								
Blood–Brain Barrier	BBB-	0.5083	BBB-	0.6045	BBB-	0.5304	BBB-	0.5113	BBB-	0.5236	BBB-	0.9967
Human Intestinal Absorption	HIA+	0.7561	HIA+	0.7564	HIA+	0.7399	HIA+	0.6005	HIA+	0.7588	HIA-	8006.0
Caco-2 Per- meability	Caco2–	0.5495	Caco2-	0.5417	Caco2–	0.5446	Caco2–	0.5571	Caco2-	0.5540	Caco2–	0.8722
P-glycoprotein Substrate	Non-substrate	0.5584	Non-substrate	0.5789	Non-substrate	0.5646	Non-substrate	0.5308	Non-substrate	0.5501	Substrate	0.5741
P-glycoprotein Inhibitor	Inhibitor Non-inhibitor	0.7905 0.8553	Inhibitor Non-inhibitor	0.6973 0.8403	Inhibitor Non-inhibitor	0.8281 0.7650	Inhibitor Non-inhibitor	0.7550 0.8230	Inhibitor Non-inhibitor	0.6992 0.8477	Non-inhibitor Non-inhibitor	0.9665 0.9968
Renal Organic Cation Transporter	Non-inhibitor	0.9243	Non-inhibitor	0.9150	Non-inhibitor	0.9317	Non-inhibitor	0.9298	Non-inhibitor	0.9264	Non-inhibitor	0.9636
Subcellular localization	Mitochondria	0.7611	Mitochondria	0.7507	Mitochondria	0.7582	Mitochondria	0.7627	Mitochondria	0.7544	Lysosome	0.4840
CYP450 2C9 Substrate	Non-substrate	0.7534	Non-substrate	0.7083	Non-substrate	0.7404	Non-substrate	0.7504	Non-substrate	0.7073	Non-substrate	0.8430
CYP450 2D6 Substrate	Non-substrate	0.8274	Non-substrate	0.8280	Non-substrate	0.8257	Non-substrate	0.8260	Non-substrate	0.8246	Non-substrate	0.8446
CYP450 3A4 Substrate	Non-substrate	0.5586	Non-substrate	0.5233	Non-substrate	0.5220	Non-substrate	0.5522	Non-substrate	0.5157	Non-substrate	0.5478
CYP450 1A2 Inhibitor	Inhibitor	0.5802	Inhibitor	0.5980	Inhibitor	0.5298	Inhibitor	0.5501	Inhibitor	0.5444	Non-inhibitor	0.9045
CYP450 2C9 Inhibitor	Inhibitor	0.6813	Inhibitor	0.6867	Inhibitor	0.7174	Inhibitor	0.6712	Inhibitor	0.6617	Non-inhibitor	0.9070
CYP450 2D6 Inhibitor	Non-inhibitor	0.8790	Non-inhibitor	0.8714	Non-inhibitor	0.8845	Non-inhibitor	0.8792	Non-inhibitor	0.8748	Non-inhibitor	0.9231
CYP450 2C19 Inhibitor	Inhibitor	0.7608	Inhibitor	0.7984	Inhibitor	0.7552	Inhibitor	0.7622	Inhibitor	0.7662	Non-inhibitor	0.9150
CYP450 3A4 Inhibitor	Non-inhibitor	0.6857	Non-inhibitor	0.6323	Non-inhibitor	0.5270	Non-inhibitor	0.6925	Non-inhibitor	0.6780	Non-inhibitor	0.8309
CYP Inhibi- tory Promis- cuity	High CYP Inhibitory Promiscuity	0.9142	High CYP Inhibitory Promiscuity	0.9426	High CYP Inhibitory Promiscuity	0.9207	High CYP Inhibitory Promiscuity	0.9196	High CYP Inhibitory Promiscuity	0.9303	Low CYP Inhibitory Promiscuity	0.9767
Human Ether-a-go-	Weak inhibitor	0.9884	Weak inhibitor	0.9803	Weak inhibitor	0.9874	Weak inhibitor	0.9953	Weak inhibitor	0.9927	Weak inhibi- tor	0.9996
go-Related Gene Inhibi-	Non-inhibitor	0.6982	Non-inhibitor	0.7508	Non-inhibitor	0.6861	Non-inhibitor	0.6717	Non-inhibitor	0.6908	Non-inhibitor	0.8761

Table 5 (continued)	nued)											
Mode	Compound 3		Compound 4		Compound 8		Compound 9		Compound 12		Amoxylin	
	Result	Probability Result	Result	Probability Result	Result	Probability Result	Result	Probability result	result	Probability Result	Result	Probability
AMES Toxic- Non AMES ity toxic	Non AMES toxic	0.6397	Non AMES toxic	0.6410	Non AMES toxic	0.6508	Non AMES toxic	0.6381	Non AMES toxic	0.6368	Non AMES toxic	0.9099
Carcinogens	Non-carcino- gens	0.7102	Non-carcino- gens	0.6973	Non-carcino- gens	0.6901	Non-carcino- gens	0.7107	Non-carcino- gens	0.6826	Non-carcino- gens	0.5439
Fish Toxicity	High FHMT	0.9961	High FHMT	0.9973	High FHMT	0.9962	High FHMT	0.9951	High FHMT	0.9962	High FHMT	0.9725
Tetrahymena Pyriformis Toxicity	High TPT	0.9914	High TPT	0.9944	High TPT	0.9947	High TPT	0.9932	High TPT	0.9951	High TPT	0.7245
Honey Bee Toxicity	Low HBT	0.7487	Low HBT	0.7543	Low HBT	0.7217	Low HBT	0.7357	Low HBT	0.7450	Low HBT	0.6935
Biodegrada- tion	Not ready bio- 1.0000 degradable	1.0000	Not ready bio- degradable	1.0000	Not ready bio- 1.0000 degradable	1.0000	Not ready bio- 1.0000 degradable	1.0000	Not ready bio- 1.0000 degradable	1.0000	Not ready biodegrad- able	0.9606

(fluconazole). Other ADMET properties were reported in Table 6.

4 Conclusion

The unceasing design and development of drugs to cure infections from bacteria and fungi increases nowadays and this has reduced the rate of mortality and morbidity globally. Therefore, study of anti-*e*-coli and anti-1,4 α -demethylase properties of 3,4-dihydropyrimidin-2(1H)-one Urea Derivatives was observed in this work using density functional theory as well as molecular docking software (Pymol, Autodock Tool, Auto dock vina and discovery studio). The descriptors which revealed the anti-*e*-coli and anti-1.4 α -demethylase were clearly identified to be E_{HOMO}, E_{LUMO}, LogP, Vol, PSA, Pol, NOR and E_{HOMO}, E_{LUMO}, LogP, Vol, Pol, NOR respectively using multiple linear regression method. The obtained molecular descriptors described well anti-e-coli and anti-1,4α-demethylase properties of 3,4-dihydropyrimidin-2(1H)-one urea derivatives. Also, compound 4 and 3 were observed to inhibit *e-coli* and 1.4α -demethylase better than other studied compounds and it was observed that every part of the studied compounds interacted with the studied proteins except the urea part of the compounds. More so, ADMET properties of five selected compounds were determined and it was observed that they were correlated to the standards used in this work. Therefore, the developed QSAR model will in help in designing and developing more efficient drug like compounds.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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Blood-Brain Barrier Human Intestinal	componing o		Compound 6		compound o		Compound 10		Compound 15		Fluconazole	
Blood-Brain Barrier Human Intestinal	Result	Probability	Result	Probability	Result	Probability	Result	Probability	Result	Probability	Result	Probability
Human Intestinal	BBB-	0.5083	BBB-	0.5113	BBB-	0.5304	BBB-	0.5236	BBB+	0.5255	BBB+	0.9382
TIONATOSON	HIA+	0.7561	HIA+	0.6005	HIA+	0.7399	HIA+	0.7588	HIA+	0.8164	HIA+	0.9894
Caco-2 Per- meability	Caco2–	0.5495	Caco2-	0.5571	Caco2–	0.5446	Caco2-	0.5540	Caco2–	0.5643	Caco2+	0.8867
P-glycoprotein Substrate	P-glycoprotein Non-substrate Substrate	0.5584	Non-substrate	0.5308	Non-substrate	0.5646	Non-substrate	0.5501	Non-substrate	0.5849	Non-substrate	0.6008
P-glycoprotein Inhibitor	Inhibitor Non-inhibitor	0.7905 0.8553	Inhibitor Non-inhibitor	0.7550 0.8230	Inhibitor Non-inhibitor	0.8281 0.7650	Inhibitor Non-inhibitor	0.6992 0.8477	Inhibitor Non-inhibitor	0.7548 0.6972	Non-inhibitor Non-inhibitor	0.8782 0.9004
Renal Organic Cation Transporter	Non-inhibitor	0.9243	Non-inhibitor	0.9298	Non-inhibitor	0.9317	Non-inhibitor	0.9264	Non-inhibitor	0.9298	Non-inhibitor	0.6461
Subcellular localization	Mitochondria	0.7611	Mitochondria	0.7627	Mitochondria	0.7582	Mitochondria	0.7544	Mitochondria	0.7408	Mitochondria	0.8498
CYP450 2C9 Substrate	Non-substrate	0.7534	Non-substrate	0.7504	Non-substrate	0.7404	Non-substrate	0.7073	Non-substrate	0.7291	Non-substrate	0.7898
CYP450 2D6 Substrate	Non-substrate	0.8274	Non-substrate	0.8260	Non-substrate	0.8257	Non-substrate	0.8246	Non-substrate	0.8249	Non-substrate	0.9116
CYP450 3A4 Substrate	Non-substrate	0.5586	Non-substrate	0.5522	Non-substrate	0.5220	Non-substrate	0.5157	Non-substrate	0.5551	Non-substrate	0.5650
CYP450 1A2 Inhibitor	Inhibitor	0.5802	Inhibitor	0.5501	Inhibitor	0.5298	Inhibitor	0.5444	Inhibitor	0.5483	Non-inhibitor	0.6312
CYP450 2C9 Inhibitor	Inhibitor	0.6813	Inhibitor	0.6712	Inhibitor	0.7174	Inhibitor	0.6617	Inhibitor	0.6493	Non-inhibitor	0.5497
CYP450 2D6 Inhibitor	Non-inhibitor	0.8790	Non-inhibitor	0.8792	Non-inhibitor	0.8845	Non-inhibitor	0.8748	Non-inhibitor	0.8862	Non-inhibitor	0.8090
CYP450 2C19 Inhibitor	Inhibitor	0.7608	Inhibitor	0.7622	Inhibitor	0.7552	Inhibitor	0.7662	Inhibitor	0.7833	Inhibitor	0.5320
CYP450 3A4 Inhibitor	Non-inhibitor	0.6857	Non-inhibitor	0.6925	Non-inhibitor	0.5270	Non-inhibitor	0.6780	Non-inhibitor	0.7530	Non-inhibitor	0.8196
CYP Inhibi- tory Promis- cuity	High CYP Inhibitory Promiscuity	0.9142	High CYP Inhibitory Promiscuity	0.9196	High CYP Inhibitory Promiscuity	0.9207	High CYP Inhibitory Promiscuity	0.9303	High CYP Inhibitory Promiscuity	0.9218	Low CYP Inhibitory Promiscuity	0.5240
Human Ether-a-go-	Weak inhibitor	0.9884	Weak inhibitor	0.9953	Weak inhibitor	0.9874	Weak inhibitor	0.9927	Weak inhibitor	0.9966	Weak inhibi- tor	0.8229
go-Related Gene Inhibi-	Non-inhibitor	0.6982	Non-inhibitor	0.6717	Non-inhibitor	0.6861	Non-inhibitor	0.6908	Non-inhibitor	0.6751	Non-inhibitor	0.6614

Mode	Compound 3		Compound 6		Compound 8		Compound 10		Compound 15		Fluconazole	
	Result	Probability Result	Result	Probability Result	Result	Probability Result	Result	Probability Result	Result	Probability Result	Result	Probability
AMES Toxic- ity	AMES Toxic- Non AMES ity toxic	0.6397	Non AMES toxic	0.6381	Non AMES toxic	0.6508	Non AMES toxic	0.6368	Non AMES toxic	0.6250	Non AMES toxic	0.5480
Carcinogens	Non-carcino- gens	0.7102	Non-carcino- gens	0.7107	Non-carcino- gens	0.6901	Non-carcino- gens	0.6826	Non-carcino- gens	0.6843	Non-carcino- gens	0.7298
Fish Toxicity	High FHMT	0.9961	High FHMT	0.9951	High FHMT	0.9962	High FHMT	0.9962	High FHMT	0.9957	High FHMT	0.7896
Tetrahymena Pyriformis Toxicity	High TPT	0.9914	High TPT	0.9932	High TPT	0.9947	High TPT	0.9951	High TPT	0.9926	High TPT	0.8590
Honey Bee Toxicity	Low HBT	0.7487	Low HBT	0.7357	Low HBT	0.7217	Low HBT	0.7450	Low HBT	0.6830	Low HBT	0.8709
Biodegrada- tion	Not ready bio- 1.0000 degradable	1.0000	Not ready bio- degradable	1.0000	Not ready bio- 1.0000 degradable	1.0000	Not ready bio- 1.0000 degradable	1.0000	Not ready bio- 1.0000 degradable	1.0000	Not ready biodegrad- able	1.0000

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